

CATALOG DOCUMENTATION
EMAP-ESTUARIES PROGRAM LEVEL DATABASE
1993 VIRGINIAN PROVINCE
BENTHIC SPECIES DATA

TABLE OF CONTENTS

1. DATA SET IDENTIFICATION
2. INVESTIGATOR INFORMATION
3. DATA SET ABSTRACT
4. OBJECTIVES AND INTRODUCTION
5. DATA ACQUISITION AND PROCESSING METHODS
6. DATA MANIPULATIONS
7. DATA DESCRIPTION
8. GEOGRAPHIC AND SPATIAL INFORMATION
9. QUALITY CONTROL/QUALITY ASSURANCE
10. DATA ACCESS
11. REFERENCES
12. TABLE OF ACRONYMS
13. PERSONNEL INFORMATION

1. DATA SET IDENTIFICATION

1.1 Title of Catalog document

EMAP-Estuaries Program Level Database
1993 Virginian Province
Benthic Taxon Data Summarized by Station

1.2 Authors of the Catalog entry

Charles Strobel, U.S. EPA NHEERL-AED
Melissa Hughes, OAO Corporation

1.3 Catalog revision date

5 April 1996

1.4 Data set name

BEN_SPEC

1.5 Task Group

Estuaries

1.6 Data set identification code

00110

1.7 Version

001

1.8 Requested Acknowledgment

If you plan to publish these data in any way, EPA requires a standard statement for work it has supported:

"Although the data described in this article have been funded wholly or in part by the U. S. Environmental Protection Agency through its EMAP-Estuaries Program, it has not been subjected to Agency review, and therefore does not necessarily reflect the views of the Agency and no official endorsement should be inferred."

2. INVESTIGATOR INFORMATION

2.1 Principal Investigator

Darryl Keith
U.S. Environmental Protection Agency
NHEERL-AED

2.2. Investigation Participant-Sample Collection

Charles J. Strobel
U.S. Environmental Protection Agency
NHEERL-AED

2.3 Principal Investigator-Sample Processing

Dr. Jeffrey B. Frithsen
Versar, Inc.

3. DATA SET ABSTRACT

3.1 Abstract of the Data Set

The BENTHIC SPECIES data set presents summary data on each benthic taxon identified across all acceptable grabs collected at a station. A count of organisms of the taxon identified from all grabs (generally 3) is recorded. The mean abundance and standard deviation of the mean abundance is also reported. Each taxon is identified by a unique code which can be cross-referenced to the taxon phylogeny. Physical constraints or quality assurance problems precluded the collection or analysis of all samples at a few stations.

3.2 Keywords for the Data Set

Benthic Species, Mean Species Abundance, Species Abundance, Species Composition, Taxon Abundance, Benthic Taxon Abundance, Mean Benthic Taxon Abundance

4. OBJECTIVES AND INTRODUCTION

4.1 Program Objective

The Environmental Monitoring and Assessment Program (EMAP) was designed to periodically estimate the status and trends of the Nation's ecological resources on a regional basis. EMAP provides a strategy to identify and bound the extent, magnitude and location of environmental degradation and improvement on a regional scale based on randomly located station sites. The randomly located stations were called Base Sampling Sites (BASE).

4.2 Data Set Objective

The objective of the Benthic Species data set is to provide summary data for each taxon or species of bottom dwelling (benthic) organism identified from each station sampled in the Virginian Province in 1993.

4.3 Data Set Background Information

Benthic invertebrates are important secondary consumers in most estuarine systems, represent the largest living reservoir of organic carbon in many estuarine systems, contain many commercially and recreationally important species and are prey for critical life stages of other commercially and recreationally important species.

Benthic invertebrate assemblages are sensitive to disturbance and stress from both natural and anthropogenic origins because of their taxonomic diversity, wide range of physiological tolerances to stress and multiple feeding modes and trophic levels. The condition of these communities is a reflection of local environmental conditions (since members of benthic assemblages generally have limited mobility). The communities respond to

both sediment and water column conditions and contain long-lived species relative to most invertebrate communities in the water column. Consequently, benthic community studies have been used in many regional estuarine monitoring programs and have proven to be an effective indicator for describing the extent and magnitude of pollution impacts in estuarine ecosystems.

Benthic monitoring data describing species composition, abundance and biomass were used as indicators of the biological conditions in the estuaries of the Virginian Province. These descriptions, along with additional measurements in other data sets describing habitat indicators (depth, salinity) and pollution exposure indicators (oxygen concentrations, sediment toxicity, sediment contaminant concentrations) are being used to develop a benthic index of environmental condition for the Province.

4.4 Summary of Investigation Parameters

Benthic species diversity, abundance and biomass were counted or measured from the grabs, generally three, collected at a station. Summary data were calculated from these laboratory data.

5. DATA ACQUISITION AND SAMPLING METHODS

5.1 Data Acquisition

5.1.1 Sampling Objective

Collect sediment grab samples suitable for the analysis of benthic assemblages and biomass. Three replicate sediment samples were expected to be taken at each station.

5.1.2 Sample Collection Methods Summary

The grab sampler was lowered through the water column such that travel through the last 5 meters was no faster than 1 m/sec. The grab penetrated the sediment by gravity releasing a trigger allowing the jaws to close. When the grab was pulled from the sediment using the winch, the jaws closed, encapsulating the sediment sample. After the sampler was retrieved, it was lowered into an on-board cradle.

5.1.3 Sampling Start Date

27 July 1993

5.1.4 Sampling End Date

31 August 1993

5.1.5 Platform

Sampling was conducted from 8 m (24 ft), twin-engine Chesapeake style work boats.

5.1.6 Sampling Gear

A 1/25 m², stainless steel, Young-modified Van Veen Grab sampler was used to collect sediment grabs for benthic analyses. This grab sampled a sample area of 440 cm² and a maximum depth of penetration in the sediment of 10 cm. Samples were sieved through a 0.5 mm round stainless steel sieve.

5.1.7 Manufacturer of Sampling Equipment

Young's Welding, Sandwich, MA

5.1.8 Key Variables

At the time of sample collection, the number of grabs collected was recorded.

5.1.9 Collection Method Calibration

The sampling gear did not require any calibration. It required inspection for deformities incurred due to mishandling or impact on rocky substrates.

5.1.10 Sample Collection Quality Control

To ensure the integrity of the sediment samples collected, the interior surfaces of the grab sampler (including the underside of the hinged top) were rinsed prior to use to assure that no sediment remained from the previous station. To minimize the effects of bow wave disturbance to surficial sediments, the speed of grab through the water column was reduced as it neared the bottom. To minimize the chance of sampling the exact same location twice, after three (3) grabs were taken, the boat was moved five (5) meters downstream by letting out the appropriate length of anchor line. Sediment grabs used for benthic samples were randomly interspersed with the grabs used for sediment chemistry/toxicity samples.

A successful grab had relatively level, intact sediment over the entire area of the grab and a sediment depth at the center of between 7-10 centimeters. Unacceptable grabs included those: substrates or grossly slumped surfaces. Grabs completely filled to the top, where the sediment was in direct contact with the hinged top, were also unacceptable.

The sieve was inspected immediately following the removal of the sample to ensure no organisms were left clinging to the sieve. Any organisms found were placed in the sample jar. The sieve was also thoroughly scrubbed with a stiff brush between samples.

5.1.11 Sample Collection Method Reference

Reifsteck, D. R., Strobel, C.J. and D. J. Keith. 1993. Environmental Monitoring and Assessment Program-Estuarines: 1993 Virginian Province Field Operations and Safety Manual. U.S. EPA NHEERL-AED, Narragansett, RI. June 1993.

5.1.12 Sample Collection Method Deviations

NA

5.2 Data Preparation and Sample Processing

5.2.1 Sample Processing Objective

Process sediment samples to accurately identify and enumerate all macrobenthic organisms found to the lowest taxonomic category which was possible.

5.2.2 Sample Processing Methods Summary

5.2.2.1 Field Summary

A clear plastic core was inserted into a random location in the grab. The sediment within the core was extruded into a "Whirl Pack" for benthic grain size analysis.

The sample was processed for benthic community analysis. Each grab

was placed separately into a frame holding a 500 um sieve. The sieve was placed into a sieve box containing water from the sampling station. The sieve was agitated to wash away sediments and leave organisms, detritus, sand particles and pebbles larger than 500 um. This method was used to minimize mechanical damage to fauna. A gentle flow of water over the sample was also acceptable.

The contents on the sieve were gently rinsed, using a funnel, into a bottle or bottles. The sieve was inspected for remaining organisms. These were removed by forceps and placed in the bottle. The volume of sample per sample jar was no more than 700 mL. 100 ml of 100% buffered, Rose Bengal stained stock formalin was added to each sample jar. A teaspoon-full of borax was added to the sample to assure saturation of the buffer, then the jar was filled to the rim with seawater to eliminate any air space (final concentration of approximately 10% formalin). The samples were gently mixed by inversion and placed in the dark. After processing each grab, the sieve was vigorously cleaned with water and a brush to prevent cross-contamination of samples.

5.2.2.2 Laboratory Summary

BENTHIC SAMPLES: The samples were washed through 500 um mesh sieves. Benthic fauna were sorted from the sediments, identified to species, if possible, and enumerated. Benthic fauna identified included those commonly termed 'macrofauna', i.e., those metazoan organisms retained by a 0.5 mm mesh sieve. 'Meiofaunal' groups were not identified or enumerated. These groups included: nematodes, ostracods, turbellarians, harpacticoid copepods and foraminifera. In addition to meiofauna, taxonomic groups having only planktonic forms were excluded from the identification process. Examples of these groups were copepods and cladocerans.

Benthic fauna were identified to the lowest practical taxonomic level. Macrobenthos were identified to species, except for the following groups: class anthozoa (class), subclass copepoda (order), phylum nemertinea (phylum), subclass ostracoda (subclass) and class turbellaria (class). For samples collected in low salinity (less than 5 ppt) water, oligochaetes and chironomids were identified to species, where possible. Above 5 ppt salinity, individuals of these groups from higher salinities were not further differentiated.

BIOMASS: Identified and counted organisms were grouped by categories of taxonomic and ecological significance to be used in biomass determinations, placed in vials and preserved.

Biomass was determined using formaldehyde dry weight. Soft-bodied organisms and those having significant inorganic body parts were treated separately. The dry weight biomass of soft-bodied organisms was directly measured after drying. However, hard-bodied organisms (e.g., bivalves, gastropods, and echinoderms) were acidified prior to measuring dry weight in order to remove calcium carbonate (bivalves >2 cm in length were shucked rather than acidified). Biomass measurements were made using an analytical balance with an accuracy of 0.1 mg. Biomass was determined as shell-free dry weight after drying to a constant weight at 60 degrees C.

In the data base, biomass data are reported along with an abundance

value (the number of organisms included in the sample). Data base records with a biomass value greater than zero but with an abundance equal to zero indicate that organism fragments were included in the sample.

SILT/CLAY: The procedure used to determine per cent silt/clay content is summarized below. The sediment sample was stirred, homogenized in a clean beaker and sieved using a 63 um mesh sieve. The fraction retained on the sieve (> 63 um) was transferred to a tared evaporating dish, dried in an oven and weighed as the sand weight. The filtrate fraction (< 63 um) was transferred to a 1 liter graduated cylinder, shaken to evenly distribute the particles and a set volume removed to a tared evaporating dish. The sample was dried and weighed as the silt/clay weight.

MOISTURE: A summary of the procedure used for the determination of moisture contents follows. The sample was brought to room temperature and homogenized in a beaker. An aliquot of wet sediment was placed in a tared evaporating dish and weighed immediately. The sample was dried and weight again.

5.2.3 Sample Processing Method Calibration

NA

5.2.4 Sample Processing Quality Control

To ensure that measurements were standardized, biomass measurements were made only after samples had been preserved for a minimum of two months.

5.2.5 Sample Processing Method Reference

U.S. EPA. 1995. Environmental Monitoring and Assessment Program (EMAP): Laboratory Methods Manual-Estuaries, Volume 1: Biological and Physical Analyses. U.S. Environmental Protection Agency, Office of Research and Development, Narragansett, RI. EPA/620/R-95/008.

5.2.6 Sample Processing Method Deviations

A change in protocol in the processing of samples for benthic biomass determination occurred. Samples collected were preserved in 10% formalin for at least 30 days, and then, for health and safety reasons, were transferred to ethanol prior to sorting.

6. DATA ANALYSIS AND MANIPULATIONS

6.1 Name of New or Modified Value

BSPECABN	Organisms of the Taxon:Total #
BSPEC_MA	Organisms of the Taxon:Mean #/Grab
BSPECSTD	Organisms of the Taxon:STD of Mean/Grab

6.2 Data Manipulation Description

Measurements on a 'per grab' basis were received from taxonomic laboratories. Values in this data set were calculated by 1) Summing replicate abundance over 'n' grabs, 2) taking the mean of the abundance across 'n' replicates and 3) generating a standard deviation based on the replicate abundances for each taxon.

6.3 Data Manipulation Examples

6.3.1 Total abundance for a taxon:

Abundance counts for a taxon were summed for all replicates collected at a station.

6.3.2 Mean and Standard Deviation (SD) values for abundance

The mean for each taxon identified at a station was calculated by summing the replicate abundances and dividing by the number of grabs collected. The SD was then calculated.

7. DATA DESCRIPTION

7.1 Description of Parameters

#	Parameter Data			Parameter	
	SAS Name	Type	Len	Format	Label
1	STA_NAME	Char	8	8.	The Station Identifier
2	VST_DATE	Num	8	YYMMDD6.	The Date the Sample was Collected
3	SPECCODE	Char	8	\$8.	EMAP Taxon Code
4	BSPECABN	Num	8	6.	Organisms of the Taxon:Total #
5	BSPEC_MA	Num	8	8.2	Organisms of the Taxon:Mean #/Grab
6	BSPECSTD	Num	8	6.2	Organisms of the Taxon:STD of Mean/Grab

7.1.6 Precision to which values are reported

Total abundance is reported as a whole number.
Mean abundance and standard deviation (SD) are reported to 2 decimal places.

7.1.7 Minimum Value in Data Set

BSPECABN	1
BSPEC_MA	0
BSPECSTD	0

7.1.7 Maximum Value in Data Set

BSPECABN	7532
BSPEC_MA	2510.67
BSPECSTD	554.88

7.2 Data Record Example

7.2.1 Column Names for Example Records

STA_NAME	VST_DATE	SPECCODE	BSPECABN	BSPEC_MA	BSPECSTD
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7.2.2 Example Data Records

OBS	STA_NAME	VST_DATE	SPECCODE	BSPECABN	BSPEC_MA	BSPECSTD
1	VA93-601	930918	ACTECANA	14	4.67	3.79
2	VA93-601	930918	ACTEPUNC	3	1.00	1.00
3	VA93-601	930918	AGLAVERR	1	0.33	0.58
4	VA93-601	930918	AMPEABDI	2	0.67	0.58
5	VA93-601	930918	AMPEABVA	2	0.67	0.58

8. GEOGRAPHIC AND SPATIAL INFORMATION

8.1 Minimum Longitude

-77 Degrees 23 Minutes 37.20 Decimal Seconds

8.2 Maximum Longitude

-70 Degrees 01 Minutes 9.00 Decimal Seconds

8.3 Minimum Latitude

36 Degrees 56 Minutes 54.00 Decimal Seconds

8.4 Maximum Latitude

42 Degrees 11 Minutes 30.00 Decimal Seconds

8.5 Name of area or region

Virginian Province

Stations were located in estuaries along the East Coast of the United States from Cape Cod, Massachusetts, to Cape Henry, Virginia, at the mouth of the Chesapeake Bay. The area includes the District of Columbia and the States of Virginia, Maryland, New Jersey, Delaware, Pennsylvania, New York, Connecticut, Rhode Island and Massachusetts.

9. QUALITY CONTROL/ QUALITY ASSURANCE

9.1 Measurement Quality Objectives

Measurement quality objectives were outlined in the Quality Assurance Project Plan (Valente and Strobel, 1993). Accuracy goals are outlined below:

Benthic Community Composition	Accuracy Goal	Precision Goal	Completion Goal
Sorting	10 %		90%
Counting	10 %		90%
Taxonomic Identification	10 %		90%
Biomass		10 %	

9.2 Quality Assurance/Control Methods

9.2.1 Sample Collection Quality Control

Following sieving, the sieve was carefully inspected to ensure that no organisms remained.

Each crew was visited during the sampling period by the QA Coordinator or Logistics coordinator. Part of the review included observing sample collection procedures to ensure samples were being processed properly.

9.2.2 Sample Processing Quality Control

Quality control for processing grab samples involves both sorting and counting check systems. A check on the efficiency of the sorting process was required to document the accuracy of the organism extraction process.

Checks on the accuracy of sample counting were conducted in conjunction with taxonomic identification and used the same criteria.

The Quality control check on each technician's efficiency at sorting (i.e., separating organisms from sediment and debris) consists of a independent re-sort by a second, experienced sorter. To pass QC, the sorter's efficiency must be at least 90%, meaning no more than 10% of the organisms in the sample were missed. A minimum of 10 percent of samples processed by a given sorter should be subjected to a QC sort at regular intervals during sample processing. If a sorter fails QC sorts, then all samples processed from the last successful QC check were resorted and any additional organisms found were added to each sample. If QC sorting passes, but some organisms were found, these animals WERE NOT added to the original sample sort.

As organisms were identified and corrected, a voucher specimen collection was compiled. This specimen collection can be used as a quality cross check by sending specimens to a separate laboratory for identification. All specimens were to be taxonomically confirmed by an outside source and any discrepancies resolved. Identification and enumeration accuracy were checked internally by a second taxonomist for at least 10 percent of the samples processed by a given technician. There should be no more than 10 percent total error (for all species) in identification or enumeration in any sample. The same procedures for sample reprocessing that are used for sorting apply to identification and counting.

Biomass determination procedures involve drying and weighing a sample. Duplicate weight measurements by a separate technician were taken before and after drying of 10 % of the samples to control and document the precision of this measurement process. If the two technicians' results differ by more than 10 percent, the source of error was identified and corrected before analysis proceeded. A series of blanks (no less than 5% of the number of samples being processed) were also included in the set of samples being dried as an additional QC check. The weight of these blanks should have varied by no more than 0.1 mg. If greater variations were found, the balance and the procedures used by the technician in its operation were checked and corrective action taken, if necessary.

9.3 Quality Assessment Results

Two QA steps were required by the EMAP-VP 1993 QA Project Plan: in-house QC checks (i.e., resorts, recounts, and ID confirmation) on 10% of each technician's work, and independent verification of species identification. The recounts (multiple types - see Table 9-3) and preliminary species verification were performed by the laboratory performing the analyses. Most of these met the requirements established in the QA Plan. Both of the laboratories performing these analyzes were evaluated by independent laboratories in 1990 or 1991; therefore, the use of such an independent evaluation in 1993 was deemed unnecessary.

Table 9-3. Results of recounts performed by the laboratory processing benthic infauna samples in 1993. Approximately 10% of all samples were processed in duplicate.

Measurement	Mean Error	Range of Error
Benthic sorting	2.9%	0 - 8.9%
Species identification and enumeration	0.75%	0 - 6.7%
Biomass	0.07%	0 - 0.8%
Weighing blanks for biomass	1.1×10^{-4} g	0 - 9×10^{-4} g

9.4 Unassessed Errors

The methods used to process benthic samples require that a small number of representative specimens of each species be set aside in a taxonomic reference collection. However, the biomass of specimens saved for the reference collection could not be measured or estimated. In most cases, specimens in the reference collection were estimated to represent a small percentage of the total macrofaunal biomass. Nonetheless, the total biomass is underestimated for those samples from which reference specimens were taken.

Total macrofaunal biomass was also potentially underestimated for samples from tidal fresh and oligohaline salinity regions where the number of chironomids or the number of oligochaetes was less than 20. Where oligochaetes and chironomids were present in sufficient numbers (>20), half were mounted on slides to complete taxonomic identifications and half were used for biomass measurements. In those instances where the number of oligochaetes or chironomids was <20 , all specimens were mounted for identification and no biomass measurements were made. This procedure generally has a negligible effect on biomass estimates.

An additional source of error results from the process of removing an aliquot of sediment from each grab for grain size analysis. This sample (a 2 cm core) was removed from each grab prior to sieving. No attempt was made to "correct" for the animals potentially lost to this sample.

10. DATA ACCESS

10.1 Data Access Procedures

Data can be downloaded from the WWW server.

10.2 Data Access Restrictions

10.3 Data Access Contact Persons

John Paul, Ph.D.
U.S. EPA NHEERL-AED
(401) 782-3037 (Tel.)
(401) 782-3030 (FAX)
paul.john@epa.gov

Data Librarian EMAP-Estuaries
U.S. EPA NHEERL-AED
(401) 782-3184 (Tel.)
(401) 782-3030 (FAX)
hughes.melissa@epa.gov

10.4 Data Set Format

Data can be downloaded in several formats from the web application and web site.

10.5 Information Concerning Anonymous FTP

Not accessible

10.6 Information Concerning WWW

Data can be downloaded from the WWW server.

10.7 EMAP CD-ROM Containing the Data Set

Data not available on CD-ROM.

11. REFERENCES

Frithsen, J.B., 1991. Technical scientific assistance to EMAP Near Coastal Group for the processing and integration of EMAP-NC Demonstration Project benthic data. Report completed 6 May 1991 for the U.S. Environmental Protection Agency, Washington, D.C. by Versar, Inc, Columbia, MD.

Holland, A.F., ed. 1990. Near Coastal Program Plan for 1990: Estuaries. EPA 600/4-90/033. U.S. EPA, NHEERL-AED, Office of Research and Development, Narragansett, RI. November 1990.

Reifsteck, D.R., Strobel, C.J. and D. J. Keith. 1993. Environmental Monitoring and Assessment Program-Estuaries: 1993 Virginian Province Field Operations and Safety Manual. U.S. EPA NHEERL-AED, Narragansett, RI. June 1993.

U.S. EPA. 1995. Environmental Monitoring and Assessment Program (EMAP): Laboratory Methods Manual-Estuaries, Volume 1: Biological and Physical Analyses. U.S. Environmental Protection Agency, Office of Research and Development, Narragansett, RI. EPA/620/R-95/008.

Valente, R. and C.J. Strobel. 1993. Environmental Monitoring and Assessment Program-Estuaries: 1993 Virginian Province Quality Assurance Project Plan. U.S. EPA NHEERL-AED, Narragansett, RI. May 1993.

12. TABLE OF ACRONYMS

13. PERSONNEL INFORMATION

Virginian Province Manager
Darryl Keith
U.S. Environmental Protection Agency
NHEERL-AED
27 Tarzwell Drive
Narragansett, RI 02882-1197
(401)782-3135 (Tel.)
(401)782-3030 (FAX)
keith.darryl@epa.gov

Virginian Province Quality Assurance Officer
Charles J. Strobel
U.S. Environmental Protection Agency
NHEERL-AED
27 Tarzwell Drive
Narragansett, RI 02882-1197
(401)782-3180 (Tel.)
(401)782-3030 (FAX)
strobel.charles@epa.gov

Principal Investigator-Sample Processing
Dr. Jeffrey B. Frithsen
Versar, Inc.
9200 Rumsey Road
Columbia, MD 21045-1934
(410)964-9200 (Tel.)
(410)964-5156 (FAX)
frithsenjef@versar.com

John Paul, Ph.D.
U.S. Environmental Protection Agency
NHEERL-AED
27 Tarzwell Drive
Narragansett, RI 02882-1197
(401) 782-3037 (Tel.)
(401) 782-3030 (FAX)
paul.john@epa.gov

Data Librarian, EMAP-Estuaries
Melissa M. Hughes
OAO Corporation
U.S. EPA NHEERL-AED
27 Tarzwell Drive
Narragansett, RI 02882-1197
(401) 782-3184 (Tel.)
(401) 782-3030 (FAX)
hughes.melissa@epa.gov